

Tail snipping and Ear Notching for Queens Medical Center SOP

First issued: 2006, Revised 7/23/09DT

Supplies:

- All procedures done under a fume hood or Biosafety cabinet
- Sharps container
- Plastic isoflurane chamber
- Isoflurane
- Gauze pads/paper towels
- Sterile surgical blades
- Microscope slides
- Quick stop
- Kim wipes
- DS-180 / Bleach
- Cooler with ice packs
- Labeled tail tubes / Tube Rack
- Ear notcher
- Tweezers
- Sample container

Procedure:

1. Arrange cages with litters of pups in order of their dates of birth.
2. Line the bottom of the Isoflurane chamber with gauze pads or paper towels. Add Isoflurane to the pads until they are completely soaked and quickly put the top on the chamber to avoid inhaling any fumes.
3. Put on gloves and have 10% bleach handy to spray your gloves in between animals to sanitize.
4. Lay down drape or paper towels. Lay down a clean Kim wipe with a clean microscope slide on top of it.
5. Open the surgical blade and allow it to drop onto the slide.
6. Males are always done first so mark the tails of the females with a marker to make it easier to identify the males and be sure not to miss any.
7. Place your first male into the chamber and wait until he is anesthetized.
8. First notch the ear of the pup with your designated ID number (numbers 1-66). You can use a pair of tweezers to remove the fragment of skin if it sticks to the ear. Then use the sterile surgical blade to remove approximately 0.5cm off the end of the tail. Use the microscope slide to cut down on.
9. Immediately place the tip of the tail in the correct labeled tube and put it on ice in the sample container in the cooler. Then return to the animal to add quick stop to the end of the tail and stop the bleeding. Return the mouse to its cage.
10. Record the animal's ear notch, gender, and tube number on a run sheet to insure that you will always be able to match that tails information to that animal.

11. Dispose of the slide and blade in the sharps container. Dispose of the Kim wipe that may be contaminated with blood in the trash. Spray your gloves with 10% bleach and dry them with a clean paper towel.
12. Check the animal to be sure it is waking up.
13. Lay down a fresh Kim wipe with a microscope slide and surgical blade. Repeat the procedure for each animal in the correct order.
14. When all mice are done, roll up your drape/paper towels and dispose of it in the trash. Wash your ear notcher and tweezers with Alconox detergent. Return all supplies to their storage location. Spray and wipe down the entire fume hood with DS-180 disinfectant
15. Seal the sample container in the cooler. A new spreadsheet must be generated to showing a map of the tubes numbers and coinciding mouse information.
16. Send an email to Queens's medical center to alert them tails are on the way and attach the spread sheet.
17. Bring the tails to the loading dock of QMC and call 537-7934 or 537-7922 or 537-7926.