

# TRPM2 Functions as a Lysosomal Ca<sup>2+</sup>-Release Channel in $\beta$ Cells

Ingo Lange,<sup>1,2</sup> Shinichiro Yamamoto,<sup>3</sup> Santiago Partida-Sanchez,<sup>4,5</sup> Yasuo Mori,<sup>3</sup> Andrea Fleig,<sup>1,2</sup> Reinhold Penner<sup>1,2\*</sup>

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TRPM2 is a Ca<sup>2+</sup>-permeable cation channel that is specifically activated by adenosine diphosphoribose (ADPR). Channel activation in the plasma membrane leads to Ca<sup>2+</sup> influx and has been linked to apoptotic mechanisms. The primary agonist, ADPR, is produced both extra- and intracellularly and causes increases in intracellular calcium concentration ([Ca<sup>2+</sup>]<sub>i</sub>), but the mechanisms involved are not understood. Using short interfering RNA and a knockout mouse, we report that TRPM2, in addition to its role as a plasma membrane channel, also functions as a Ca<sup>2+</sup>-release channel activated by intracellular ADPR in a lysosomal compartment. We show that both functions of TRPM2 are critically linked to hydrogen peroxide-induced  $\beta$  cell death. Additionally, extracellular ADPR production by the ectoenzyme CD38 from its substrates NAD<sup>+</sup> (nicotinamide adenine dinucleotide) or cADPR causes IP<sub>3</sub>-dependent Ca<sup>2+</sup> release via P2Y and adenosine receptors. Thus, ADPR and TRPM2 represent multimodal signaling elements regulating Ca<sup>2+</sup> mobilization in  $\beta$  cells through membrane depolarization, Ca<sup>2+</sup> influx, and release of Ca<sup>2+</sup> from intracellular stores.

## INTRODUCTION

TRPM2 (transient receptor potential channel, melastatin subfamily type 2) is a nonselective, Ca<sup>2+</sup>-permeable cation channel with unique gating properties that are conferred by a functional adenosine diphosphoribose (ADPR) hydrolase domain in its C terminus (1–3). TRPM2 is synergistically activated and regulated by multiple signaling pathways through various adenine dinucleotides [ADPR, cyclic ADPR (cADPR), and nicotinic acid adenine dinucleotide phosphate (NAADP)] and intracellular calcium concentration ([Ca<sup>2+</sup>]<sub>i</sub>) (1, 2, 4–9), agonists that vary in structure as well as in source. Although all of these molecules function intracellularly to activate TRPM2, some of them, in particular the primary TRPM2 agonist ADPR, can be produced extracellularly through the multifunctional ectoenzyme CD38 (10). Intracellularly, ADPR can be released from mitochondria (6) or through production of free ADPR during extreme DNA damage through the poly(ADP-ribose) polymerase (PARP)–poly(ADP-ribose) glycohydrolase (PARG) pathway (11). Moreover, apoptotic stimuli that mediate oxidative stress, such as hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) or tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), have emerged as (patho)physiological stimuli for TRPM2-mediated signaling, mediating Ca<sup>2+</sup> entry in Jurkat T cells, neutrophils, microglia, and pancreatic  $\beta$  cells (12). When applied extracellularly, the primary cytosolic agonist ADPR can also elicit Ca<sup>2+</sup> release in cells that express TRPM2. This mechanism is not well understood, but has been suggested to result from inositol 1,4,5-trisphosphate (IP<sub>3</sub>) production (13). Here, we examine the signaling mechanisms of ADPR in heterologous expression systems and in natively TRPM2-expressing pancreatic  $\beta$  cells, where we find that TRPM2 serves a dual role as plasma membrane Ca<sup>2+</sup>-influx channel and as a pre-

viously unidentified intracellular Ca<sup>2+</sup>-release channel, with both functions playing a critical role in H<sub>2</sub>O<sub>2</sub>-induced  $\beta$  cell death.

## RESULTS

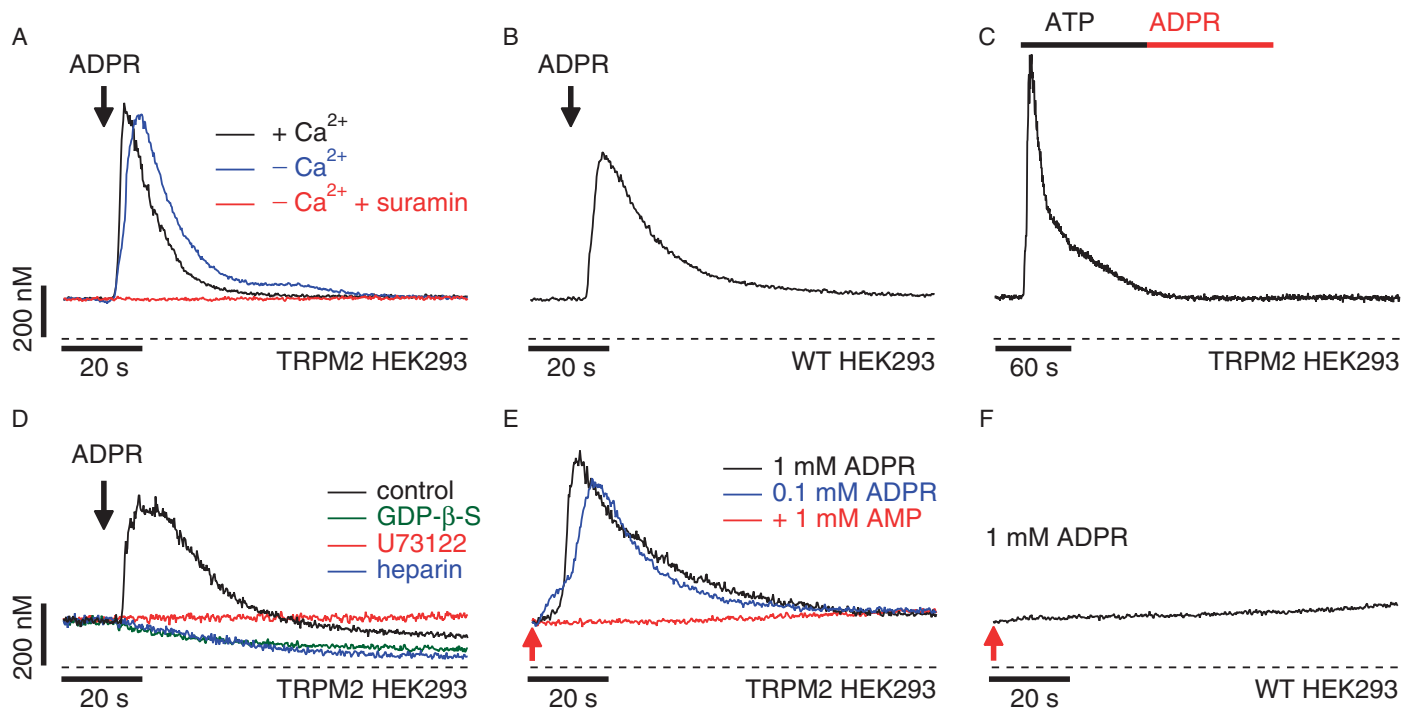
### Extracellular ADPR activates P2Y receptors in HEK293 cells

Extracellular nucleosides and different ribosylated nucleotide derivatives have been shown to activate Ca<sup>2+</sup> signaling pathways through IP<sub>3</sub>-producing receptors (13). Thus, before we investigated the possible effects of intracellular ADPR on Ca<sup>2+</sup> release from internal stores, we first explored the signaling function of extracellular ADPR. Wild-type human embryonic kidney (HEK) 293 cells do not express native TRPM2 (1). Nevertheless, addition of ADPR to intact HEK293 cells consistently produced a transient Ca<sup>2+</sup> signal in both wild-type cells (Fig. 1B) and cells heterologously expressing TRPM2 (TRPM2 HEK293; Fig. 1A). The Ca<sup>2+</sup> signal was elicited at a threshold concentration of ~100  $\mu$ M ADPR and required neither extracellular Ca<sup>2+</sup> (Fig. 1A) nor TRPM2 expression (Fig. 1B), indicating that it arose from release of Ca<sup>2+</sup> from intracellular stores through a preexisting signaling pathway that is independent of TRPM2. Consistent with this notion, emptying intracellular stores with thapsigargin before the ADPR challenge resulted in complete loss of ADPR-mediated Ca<sup>2+</sup> release (fig. S1).

In HEK293 cells, adenosine 5'-triphosphate (ATP) causes IP<sub>3</sub>-dependent Ca<sup>2+</sup> release through a phospholipase C (PLC) signaling pathway mediated by P2Y-type purinergic receptors (14–16). When TRPM2-expressing cells were preincubated for 15 min with 100  $\mu$ M suramin, a nonselective P2Y receptor antagonist, ADPR-induced Ca<sup>2+</sup> signals were completely suppressed (Fig. 1A). When HEK293 cells were sequentially exposed to ATP (100  $\mu$ M) and then to ADPR, the ATP-induced Ca<sup>2+</sup> release abrogated a subsequent Ca<sup>2+</sup> signal by ADPR (Fig. 1C). Thus, ADPR appears to engage the PLC signaling pathway. We tested this possibility by inhibiting the heterotrimeric guanine nucleotide-binding protein (G protein)–PLC–IP<sub>3</sub> signal transduction pathway in patch-clamp experiments (Fig. 1D). Interfering with G-protein coupling by internally perfusing HEK293 cells with 500  $\mu$ M guanosine 5'-O-(2'-thiodiphosphate)

<sup>1</sup>Center for Biomedical Research, The Queen's Medical Center, Honolulu, HI 96813, USA. <sup>2</sup>John A. Burns School of Medicine at the University of Hawaii, Honolulu, HI 96813, USA. <sup>3</sup>Department of Synthetic Chemistry and Biological Chemistry, Graduate School of Engineering, Kyoto University, Kyoto 615-8510, Japan. <sup>4</sup>The Research Institute at Nationwide Children's Hospital, Columbus, OH 43205, USA. <sup>5</sup>The Ohio State University College of Medicine, Columbus, OH 43205, USA.

\*To whom correspondence should be addressed. E-mail: rpenner@hawaii.edu



**Fig. 1.** ADPR acts as a purinergic receptor agonist and TRPM2 acts as a  $\text{Ca}^{2+}$ -release channel when heterologously expressed in HEK293 cells. (A) Average  $\text{Ca}^{2+}$  signals measured in intact HEK293 cells heterologously expressing TRPM2 channels (TRPM2 HEK293) in response to application of extracellular ADPR in the presence (1 mM, black trace,  $n = 8$ ) or absence (blue trace,  $n = 7$ ) of extracellular  $\text{Ca}^{2+}$  in the external solution. The concentration of ADPR was 1 mM in the presence of  $\text{Ca}^{2+}$  and 100  $\mu\text{M}$  in the absence of  $\text{Ca}^{2+}$ . The red trace represents the average  $\text{Ca}^{2+}$  signal measured in response to application of 100  $\mu\text{M}$  ADPR in the absence of extracellular  $\text{Ca}^{2+}$  with 100  $\mu\text{M}$  suramin ( $n = 6$ ). Application started as indicated by the arrow and was maintained throughout the experiment. Cells were loaded with 5  $\mu\text{M}$  fura-2-AM at 37°C for 30 min. (B) Average  $\text{Ca}^{2+}$  signals in intact wild-type HEK293 cells in response to application of 1 mM ADPR (black trace,  $n = 7$ ) in the absence of extracellular  $\text{Ca}^{2+}$ . Application and fura-2-AM loading as described in (A). (C) Average  $\text{Ca}^{2+}$  signal measured in intact fura-2-AM-loaded TRPM2 HEK293 cells in response to application of 100  $\mu\text{M}$  ATP (black bar) followed by application of 100  $\mu\text{M}$  ADPR (red bar) in the absence of extracellular  $\text{Ca}^{2+}$  ( $n = 6$ ). (D) The traces depict balanced

(GDP- $\beta$ -S) blocked ADPR-induced  $\text{Ca}^{2+}$  release. Likewise, ADPR-mediated  $\text{Ca}^{2+}$  release was eliminated by preincubation of cells with the PLC inhibitor U73122 (10  $\mu\text{M}$ ). Lastly, direct block of  $\text{IP}_3$  receptors with intracellular heparin (100  $\mu\text{g}/\text{ml}$ ) also prevented ADPR-induced  $\text{Ca}^{2+}$  release. These data indicate that ADPR can act as a first messenger through G protein-coupled P2Y receptors that activate the PLC signaling pathway.

### Intracellular ADPR elicits $\text{Ca}^{2+}$ release in TRPM2-expressing HEK293 cells

We next investigated whether intracellular ADPR can act as second messenger to mediate  $\text{Ca}^{2+}$  release from intracellular stores. We eliminated ADPR-mediated  $\text{Ca}^{2+}$  release through the P2Y pathway and  $\text{Ca}^{2+}$  influx through TRPM2 channels, respectively, by adding suramin to and removing  $\text{Ca}^{2+}$  from the extracellular medium. Cells were loaded with fura-2 acetoxy-

fura-2 experiments, in which TRPM2 HEK293 cells were preloaded with fura-2-AM and the patch pipette contained 200  $\mu\text{M}$  fura-2 to enable continuous measurements of  $[\text{Ca}^{2+}]_i$ . Whole-cell break-in was just before application of 100  $\mu\text{M}$  ADPR in the absence of extracellular  $\text{Ca}^{2+}$  as indicated by the arrow (black trace,  $n = 4$ ). The internal solution was supplemented with either heparin (100  $\mu\text{g}/\text{ml}$ ; blue trace,  $n = 6$ ) or 500  $\mu\text{M}$  GDP- $\beta$ -S (green trace,  $n = 5$ ). The red trace represents  $\text{Ca}^{2+}$  measurements in intact cells exposed to 10  $\mu\text{M}$  U73122 in the bath ( $n = 5$ ). (E) Balanced fura-2 experiments with internal perfusion of ADPR. Average  $\text{Ca}^{2+}$  signal in whole-cell patch-clamped TRPM2 HEK293 cells preloaded with fura-2-AM. Whole-cell break-in was at the time indicated by the red arrow. Cells were kept in 0  $\text{Ca}^{2+}$  external solution and perfused with internal solution containing 200  $\mu\text{M}$  fura-2 and supplemented with either 1 mM ADPR (black trace,  $n = 6$ ), 100  $\mu\text{M}$  ADPR (blue trace,  $n = 7$ ), or 100  $\mu\text{M}$  ADPR and 1 mM AMP (red trace,  $n = 8$ ). (F) Balanced fura-2 experiments in wild-type HEK293 cells preloaded with fura-2-AM. Whole-cell break-in was achieved at the time indicated by the red arrow. Internal solution contained 200  $\mu\text{M}$  fura-2 supplemented with 1 mM ADPR ( $n = 6$ ).

methyl ester (fura-2-AM) and then patch-clamped in the whole-cell configuration to introduce 0.1 to 1 mM ADPR intracellularly. Patch pipettes also contained 200  $\mu\text{M}$  fura-2 to maintain the ability to measure  $[\text{Ca}^{2+}]_i$  during whole-cell recording. TRPM2 HEK293 internally perfused with 100  $\mu\text{M}$  or 1 mM ADPR responded with  $\text{Ca}^{2+}$  release signals that were inhibited by intracellular adenosine monophosphate (AMP) (Fig. 1E), an established inhibitor of ADPR-gated TRPM2 channels (5). However, ADPR failed to elicit  $\text{Ca}^{2+}$  signals in wild-type cells (Fig. 1F), suggesting that ADPR induced  $\text{Ca}^{2+}$  release through TRPM2 channels located in intracellular stores.

### TRPM2 is natively expressed in INS-1 $\beta$ cells

Because TRPM2-like currents have been reported in rat RINm5F and CRI-G1  $\beta$  cell lines (12), we extended our investigation to the rat  $\beta$  pan-

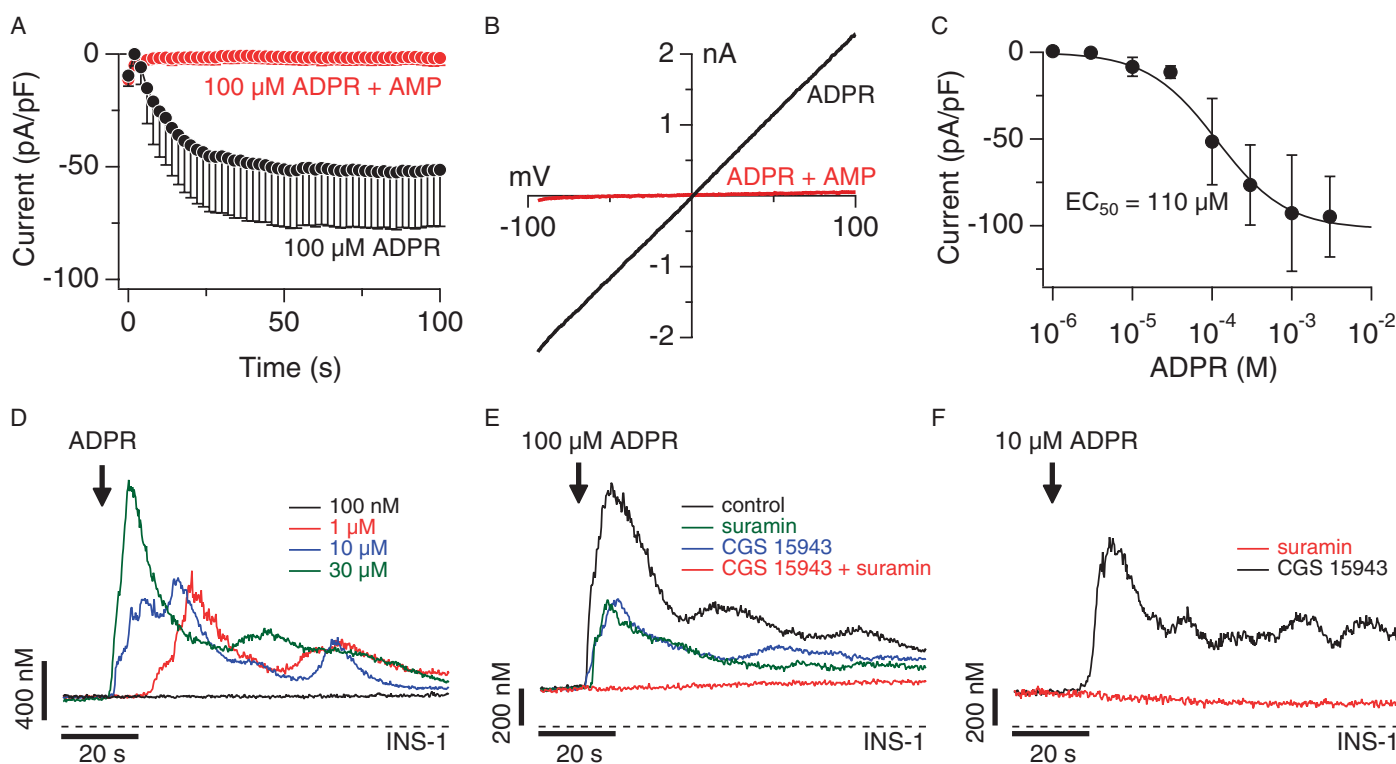
creatic cell line INS-1. Internal perfusion of INS-1 cells with ADPR caused rapid activation (Fig. 2A, open circles) of a linear current with biophysical and pharmacological characteristics typical of TRPM2 (Fig. 2B). These ADPR-mediated currents were activated in a concentration-dependent manner with a half-maximal effective concentration ( $EC_{50}$ ) of  $\sim 100 \mu\text{M}$  ADPR (Fig. 2C) and were completely suppressed by 1 mM AMP (Figs. 2A, closed circles, and 2B, red trace). These data confirm that TRPM2 is functionally expressed in INS-1 cells and acts as a plasma membrane ion channel.

### Extracellular ADPR activates P2Y and adenosine receptors in INS-1 $\beta$ cells

Before addressing the possibility that intracellular ADPR induced  $\text{Ca}^{2+}$  release in INS-1 cells, we first assessed the  $\text{Ca}^{2+}$  signaling mechanisms of extracellular ADPR in these cells. As in HEK293 cells, application of extracellular ADPR in  $\text{Ca}^{2+}$ -free solution elicited  $\text{Ca}^{2+}$  release in INS-1 cells,

but compared to HEK293 cells, at a much lower threshold concentration of  $1 \mu\text{M}$  (Fig. 2D). Although the P2Y antagonist suramin reduced the  $100 \mu\text{M}$  ADPR-induced  $\text{Ca}^{2+}$  signal, it did not abolish it (Fig. 2E). This indicated that there was another receptor type responsive to ADPR.  $\beta$  cells also express A-1 adenosine receptors (17–19), which might account for the suramin-resistant  $\text{Ca}^{2+}$  release with ADPR. We confirmed this possibility by using the broadly acting adenosine receptor antagonist CGS-15943. Like suramin, CGS-15943 reduced the ADPR-mediated  $\text{Ca}^{2+}$  signal without abolishing it (Fig. 2E). However, the combination of CGS-15943 and suramin completely suppressed the ADPR-mediated  $\text{Ca}^{2+}$  release signal (Fig. 2E). Because both P2Y and adenosine receptors can stimulate the classical G protein-coupled receptor–G protein–PLC–IP<sub>3</sub> pathway (20, 21), it is likely that the responses to extracellular ADPR in INS-1 cells are mediated by IP<sub>3</sub>-induced  $\text{Ca}^{2+}$  release.

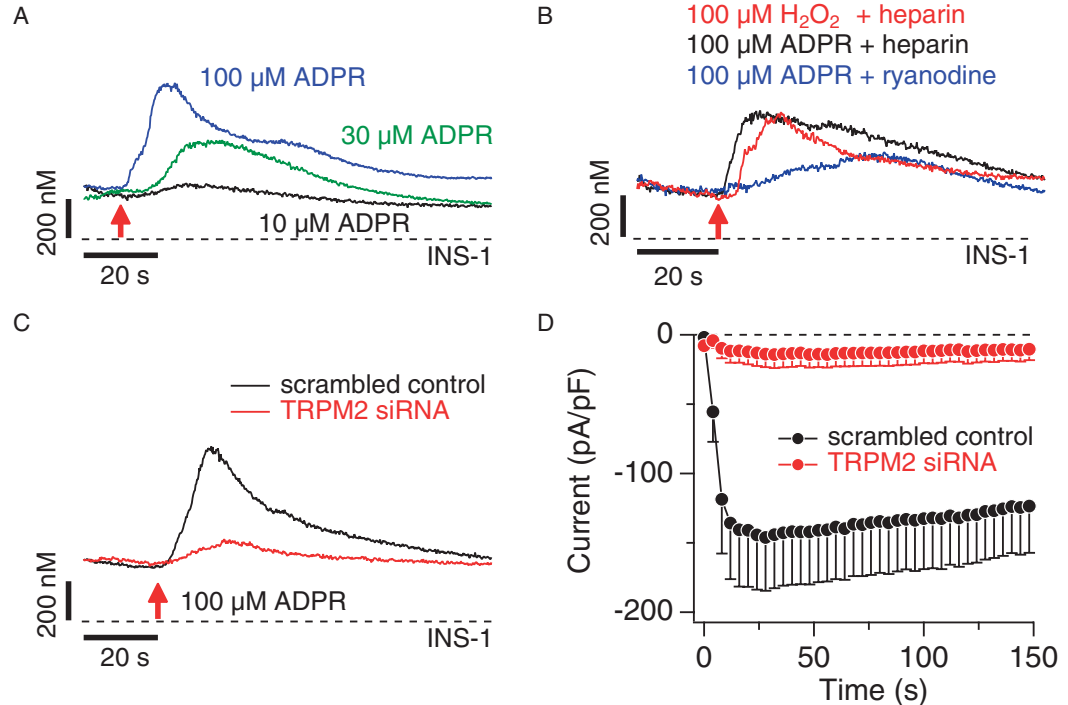
We next examined whether the enhanced ADPR sensitivity of INS-1 cells compared to HEK293 cells was mediated through adenosine or P2Y receptors by stimulating cells with  $10 \mu\text{M}$  ADPR in the presence of CGS-15943 or



**Fig. 2.** ADPR activates TRPM2, purinergic receptors, and adenosine receptors in INS-1  $\beta$  cells. (A) Average development of TRPM2 currents assessed by whole-cell patch-clamp measurements in INS-1 cells. Cells were internally perfused with either  $100 \mu\text{M}$  ADPR (black symbols,  $n = 11$ ) or  $100 \mu\text{M}$  ADPR + 1 mM AMP (red symbols,  $n = 9$ ). Current amplitudes were assessed at  $-80 \text{ mV}$ , normalized for cell size, averaged and plotted versus time of the experiment. The standard voltage protocol was ramping from  $-100 \text{ mV}$  to  $+100 \text{ mV}$  over 50 ms and at 0.5 Hz. Holding potential was 0 mV. Error bars indicate SEM. (B) Typical current-voltage ( $I$ - $V$ ) relationship of currents evoked by 1 mM ADPR (black trace), or  $100 \mu\text{M}$  ADPR + 1 mM AMP (red trace) taken from example cells and recorded 100 s into the experiment. (C) Dose-response behavior of TRPM2 currents in INS-1 cells at various internal ADPR concentrations. Current amplitudes were measured at  $-80 \text{ mV}$ , averaged, normalized to cell size, and plotted against the re-

spective ADPR concentration ( $n = 5$  to 11). A dose-response fit to the data yielded an  $EC_{50}$  value of  $110 \mu\text{M}$  with a Hill coefficient of 1. (D) Average  $\text{Ca}^{2+}$  signals measured in intact fura-2-AM-loaded INS-1 cells in response to increasing concentrations of extracellular ADPR applied in the absence of extracellular  $\text{Ca}^{2+}$  [ $100 \text{ nM}$  (black trace,  $n = 6$ ),  $1 \mu\text{M}$  (red trace,  $n = 6$ ),  $10 \mu\text{M}$  (blue trace,  $n = 6$ ),  $30 \mu\text{M}$  (green trace,  $n = 6$ )]. (E) Average  $\text{Ca}^{2+}$  signals measured in intact fura-2-AM-loaded INS-1 cells in the absence of extracellular  $\text{Ca}^{2+}$  and stimulated by  $30 \mu\text{M}$  extracellular ADPR (black trace, control,  $n = 11$ ) or  $100 \mu\text{M}$  ADPR plus either  $100 \mu\text{M}$  suramin (green trace,  $n = 8$ ) or  $1 \mu\text{M}$  CGS-15943 (blue trace,  $n = 11$ ) or both  $100 \mu\text{M}$  suramin and  $1 \mu\text{M}$  CGS-15943 (red trace,  $n = 6$ ). (F) Average  $\text{Ca}^{2+}$  signals measured in intact fura-2-AM-loaded INS-1 cells in the absence of extracellular  $\text{Ca}^{2+}$  and stimulated by  $10 \mu\text{M}$  ADPR plus either  $100 \mu\text{M}$  suramin (black trace,  $n = 6$ ) or  $1 \mu\text{M}$  CGS-15943 (red trace,  $n = 6$ ).

**Fig. 3. TRPM2 functions as  $\text{Ca}^{2+}$ -release channel in INS-1 cells.** (A) Balanced fura-2 experiments showing average  $\text{Ca}^{2+}$  signals in whole-cell patch-clamped INS-1 cells preloaded with fura-2-AM. Whole-cell break-in was at the time indicated by the red arrow. Cells were kept in  $0 \text{ Ca}^{2+}$  external solution supplemented with  $1 \mu\text{M}$  CGS-15943 and  $100 \mu\text{M}$  suramin and perfused with internal solution containing  $200 \mu\text{M}$  fura-2 and supplemented with either  $100 \mu\text{M}$  ADPR (blue trace,  $n = 9$ ),  $30 \mu\text{M}$  ADPR (green trace,  $n = 8$ ), or  $10 \mu\text{M}$  ADPR (black trace,  $n = 6$ ). (B) Balanced fura-2 experiments, showing average  $\text{Ca}^{2+}$  signals in whole-cell patch-clamped INS-1 cells preloaded with fura-2-AM. Whole-cell break-in was at the time indicated by the red arrow. Cells were kept in  $0 \text{ Ca}^{2+}$  external solution containing  $1 \mu\text{M}$  CGS-15943 and  $100 \mu\text{M}$  suramin. Cells were perfused with internal solution containing  $200 \mu\text{M}$  fura-2 and supplemented with either  $100 \mu\text{M}$   $\text{H}_2\text{O}_2$  plus heparin ( $100 \mu\text{g/ml}$ ; red trace,  $n = 10$ ),  $100 \mu\text{M}$  ADPR plus heparin ( $100 \mu\text{g/ml}$ ; black trace,  $n = 7$ ), or  $100 \mu\text{M}$  ADPR with  $25 \mu\text{M}$  external ryanodine (blue trace,  $n = 6$ ). (C) Balanced fura-2 experiments showing average  $\text{Ca}^{2+}$  signals in response to internal ADPR in whole-cell patch-clamped INS-1 cells preloaded with fura-2-AM. Whole-cell break-in was at the time indicated by the red arrow. Cells were kept in  $0 \text{ Ca}^{2+}$  external solution supplemented with  $100$



suramin. Suramin was considerably more effective than CGS-15943 in suppressing the response to the low concentration of ADPR (Fig. 2F), suggesting that P2Y receptors are primarily responsible for the higher sensitivity of INS-1 cells.

### Intracellular ADPR elicits $\text{Ca}^{2+}$ release in INS-1 $\beta$ cells

We next investigated the possibility that native TRPM2 channels in INS-1 cells mediate intracellular  $\text{Ca}^{2+}$  release. Intracellular perfusion of cells with ADPR in the absence of extracellular  $\text{Ca}^{2+}$  and with both suramin and CGS-15943 in the bath produced a concentration-dependent increase in  $[\text{Ca}^{2+}]_i$  (Fig. 3A). Because TRPM2 is a downstream target of reactive oxygen species (ROS) (1), we tested whether  $\text{H}_2\text{O}_2$  can also mediate  $\text{Ca}^{2+}$  release in these cells. Perfusing cells with  $100 \mu\text{M}$   $\text{H}_2\text{O}_2$  (plus heparin to inhibit  $\text{IP}_3$  receptors) indeed evoked  $\text{Ca}^{2+}$  release (Fig. 3B). Furthermore,  $\text{Ca}^{2+}$  release induced by direct ADPR perfusion was not prevented by inhibiting  $\text{IP}_3$  receptors with heparin ( $100 \mu\text{g/ml}$ ), or ryanodine receptors with  $25 \mu\text{M}$  ryanodine (Fig. 3B). We confirmed that the ADPR-mediated responses involved TRPM2 by molecular knockdown of TRPM2 with short interfering RNA (siRNA). TRPM2-specific siRNA, but not a scrambled control siRNA, caused a significant suppression of ADPR-induced  $\text{Ca}^{2+}$  release with  $P < 0.03$ , as assessed by peak amplitude changes in  $\text{Ca}^{2+}$  release (Fig. 3C and table S1). We confirmed the efficacy of specific siRNA knockdown of TRPM2 by monitoring TRPM2 channel activity in the plasma membrane and observed nearly complete suppression of functional channel activity (Fig. 3D). Together, these data show that TRPM2

$\mu\text{M}$  suramin and  $1 \mu\text{M}$  CGS-15943 and perfused with internal solution containing  $200 \mu\text{M}$  fura-2 and supplemented with  $100 \mu\text{M}$  ADPR. Traces represent  $\text{Ca}^{2+}$  signals from cells treated with scrambled control siRNA (black trace,  $n = 10$ ) or TRPM2-specific siRNA (red trace,  $n = 10$ ). (D) Average TRPM2 currents assessed by whole-cell patch-clamp measurements in INS-1 cells treated with scrambled control siRNA (black symbols,  $n = 8$ ) or TRPM2-specific siRNA (red symbols,  $n = 14$ ). Currents were analyzed as described in Fig. 2A.

proteins in INS-1  $\beta$  cells function as both  $\text{Ca}^{2+}$ -permeable cation channels in the plasma membrane and as  $\text{Ca}^{2+}$ -release channels in intracellular stores.

### TRPM2 is a lysosomal $\text{Ca}^{2+}$ -release channel in INS-1 $\beta$ cells

We next assessed the subcellular localization of TRPM2 in INS-1 cells by immunofluorescence and observed both peripheral and intracellular localization of TRPM2 (Fig. 4A). These data revealed that TRPM2 rarely, if ever, colocalized with the endoplasmic reticulum (ER). Instead, TRPM2 showed a punctate distribution throughout the cytoplasm, indicating localization in a vesicular compartment. Because lysosomal organelles contain  $\text{Ca}^{2+}$  and have been implicated in  $\text{Ca}^{2+}$  release (22, 23), we investigated the distribution of both TRPM2 and lysosomes in INS-1 cells with specific antibodies against TRPM2 and lysosome-associated membrane protein-1 (LAMP-1), a specific marker for lysosomes (24). Confocal images of INS-1 cells revealed dense regions of punctate labeling for both proteins that exhibited a high degree of overlap (Fig. 4B), although a few vesicular structures were labeled by only TRPM2. We assessed this store functionally by using bafilomycin A, a macrolide antibiotic that selectively inhibits the vacuolar  $\text{H}^+$ -dependent adenosine triphosphatase at nanomolar concentrations and empties lysosomal  $\text{Ca}^{2+}$  stores without affecting ER  $\text{Ca}^{2+}$  concentrations (22, 25). Indeed, preincubation of cells with  $100 \text{ nM}$  bafilomycin A for 30 min. completely suppressed  $\text{Ca}^{2+}$  release by  $300 \mu\text{M}$  ADPR subsequently introduced through the patch pipette



(Fig. 4C). Control experiments showed that IP<sub>3</sub>-mediated Ca<sup>2+</sup> release from the ER of these cells after stimulation of muscarinic receptors with 300 μM carbamylcholine was only slightly reduced by bafilomycin A pretreatment (Fig. 4D). Together, these data indicate that ADPR-dependent TRPM2-mediated Ca<sup>2+</sup> release occurs predominantly from a lysosomal store.

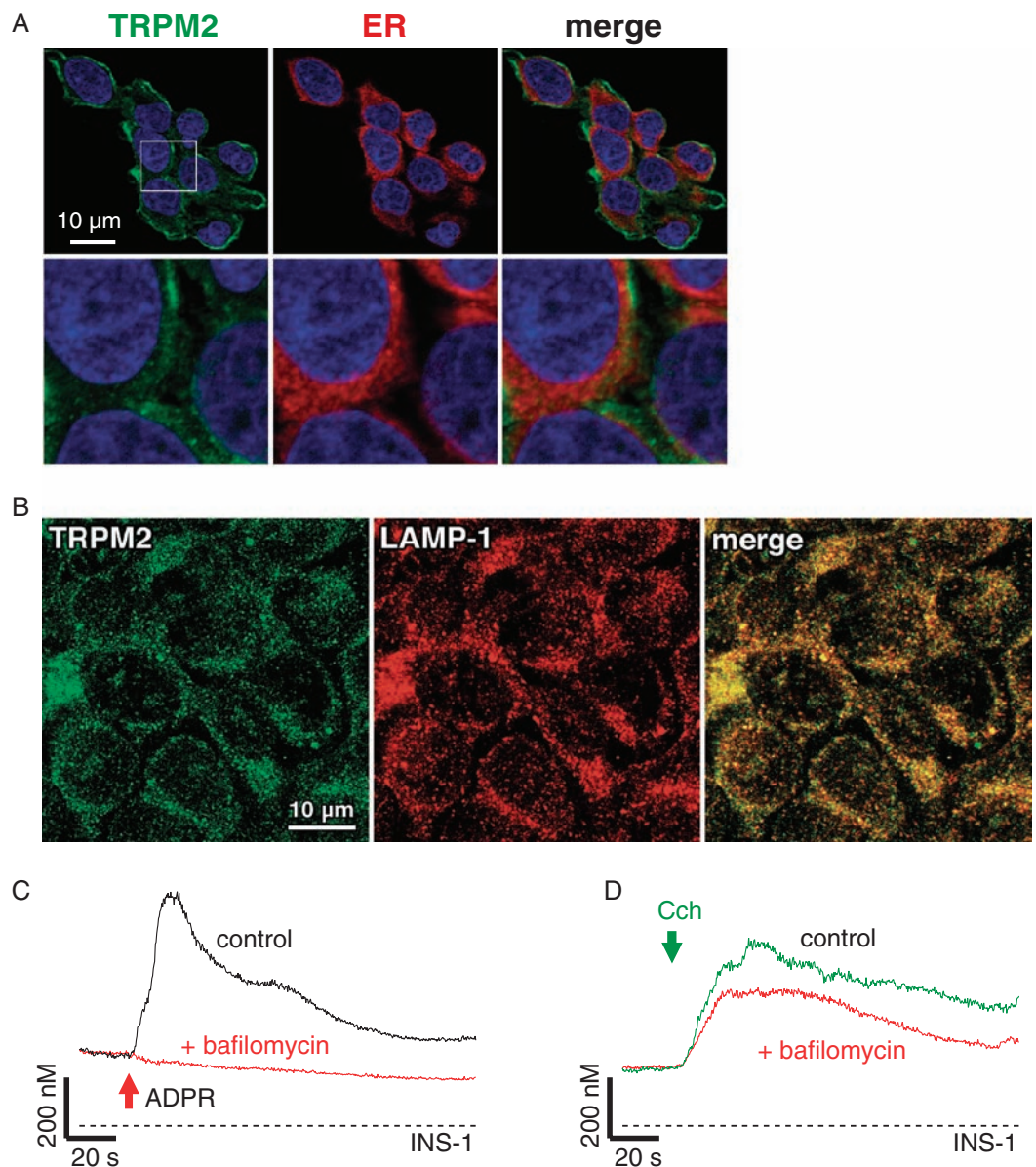
### TRPM2 is natively expressed in primary mouse β cells

Although the INS-1 cell line represents a widely used model for pancreatic β cells, cell lines do not fully reflect the properties of primary cells. We

therefore extended our analysis to primary pancreatic β cells isolated from C57BL/6 mice. First, we evaluated the potency of ADPR at activating TRPM2-like currents in the plasma membrane. Experiments were performed 24 to 72 hours after isolation of pancreatic β cells. Cells were maintained under the same conditions as the INS-1 cell line and subjected to the same experimental protocols with identical ionic composition of internal and external solutions. We internally perfused cells with various concentrations of ADPR and observed rapid activation of linear currents with biophysical and pharmacological characteristics typical of TRPM2 (Fig. 5B).

Fig. 4. TRPM2 is a lysosomal Ca<sup>2+</sup>-

release channel in INS-1 cells. (A) Detection and cellular localization of TRPM2 by immunofluorescence. Polyclonal antibodies directed against mouse TRPM2 specifically recognizes a protein in INS-1 cells with cytosolic, as well as plasma membrane distribution (left panels, green). Intracellular TRPM2 label is largely excluded from the ER (middle panels, red) network, as evidenced by the merged image (right panels, note absence of yellow spots). DAPI (4',6-diamidino-2-phenylindole) was used as a nuclear counterstain (blue). Images of cells that are representative of the entire population are shown (63× magnification). The white rectangle indicates the area of expanded view depicted in the respective lower panels. (B) Immunofluorescence of TRPM2 (left panel, green) and LAMP-1 (middle panel, red) with antibodies directed against TRPM2 (3) and LAMP-1. The right panel represents the merged image, suggesting that both proteins have largely overlapping localizations (yellow), with just a few vesicles showing only TRPM2 fluorescence. (C) Bafilomycin A inhibits intracellular ADPR-mediated Ca<sup>2+</sup> release. Balanced fura-2 experiments showing average Ca<sup>2+</sup> signals in whole-cell patch-clamped INS-1 cells preloaded with fura-2-AM. Whole-cell break-in was at the time indicated by the red arrow. Cells were kept in 0 Ca<sup>2+</sup> external solution containing 100 μM suramin and 1 μM CGS-15943 in the absence (control, black trace, *n* = 9) or presence of 100 nM bafilomycin A (red trace, *n* = 16) and perfused with internal solution containing 100 μM ADPR or 300 μM ADPR, respectively. (D) Average Ca<sup>2+</sup> signals measured in intact



intact fura-2-AM-loaded INS-1 cells in response to 300 μM carbamylcholine (CCh) in the absence of extracellular Ca<sup>2+</sup> and in the presence of 100 μM suramin and 1 μM CGS-15943 (control, black trace, *n* = 20) in the external solution. The red trace (*n* = 17) represents cells treated identically, but preincubated with 100 nM bafilomycin A for 30 min.

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These currents reached peak amplitudes of about  $-80$  pA/pF at  $-80$  mV (Fig. 5C) within 30 to 50 s and could be suppressed by AMP (Fig. 5A). The ADPR-induced currents were concentration dependent, with an  $EC_{50}$  of  $\sim 360$   $\mu$ M ADPR (Fig. 5C). Thus, TRPM2 is expressed as a functional ion channel in primary mouse  $\beta$  cells.

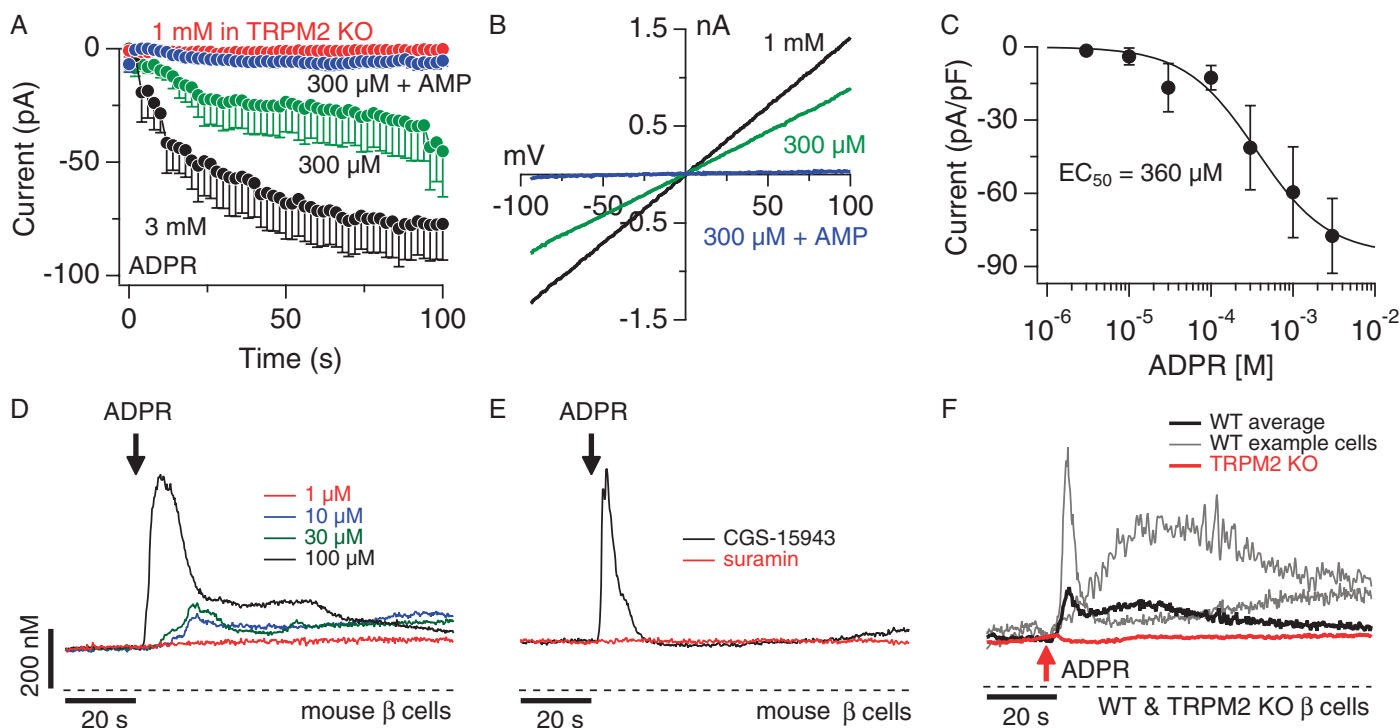
### Extracellular ADPR activates P2Y receptors in primary mouse $\beta$ cells

Before investigating TRPM2-dependent  $Ca^{2+}$  release, we assessed the extracellular effects of ADPR in primary  $\beta$  cells. With a threshold concentration of  $\sim 10$   $\mu$ M, ADPR produced multiple  $Ca^{2+}$ -release transients when applied to intact cells (Fig. 5D). Although the averaged  $Ca^{2+}$  signal obscures the oscillatory pattern of  $Ca^{2+}$  release in individual cells, a more quantitative analysis of  $Ca^{2+}$ -release signals at the single-cell level over 100 s, with a fitting routine based on a convolution of a Gaussian peak and exponential decay (Supplementary Materials and Methods and table S1), revealed that application of

extracellular ADPR induced two to three  $Ca^{2+}$  oscillations in both INS-1 and primary  $\beta$  cells (fig. S4A). Increasing ADPR concentrations both shortened the delay (fig. S4B) and increased the peak of the first  $Ca^{2+}$  transient (fig. S4C). CGS-15943 had no effect on the response, and suramin alone at 100  $\mu$ M completely abolished extracellularly mediated ADPR effects (Fig. 5E), suggesting that primary mouse  $\beta$  cells lack adenosine receptors and that  $Ca^{2+}$  signals in these cells are mediated by P2Y receptors.

### $Ca^{2+}$ release by extracellular cADPR and $NAD^+$ requires CD38 in primary mouse $\beta$ cells

Extracellular production of ADPR is mediated by the ectoenzyme CD38 with  $NAD^+$  (the oxidized form of nicotinamide adenine dinucleotide) or cADPR as substrate (10). Nevertheless, in contrast to ADPR, even high millimolar concentrations of these compounds were ineffective at producing  $Ca^{2+}$  signals in HEK293 (Fig. 6A), indicating that neither  $NAD^+$  nor cADPR act as P2Y receptor agonists in these cells. In contrast, both



**Fig. 5.** ADPR activates purinergic receptors and elicits  $Ca^{2+}$  influx, as well as  $Ca^{2+}$  release through TRPM2 in mouse pancreatic  $\beta$  cells. **(A)** Average TRPM2 currents in mouse pancreatic  $\beta$  cells isolated from C57BL/6 or TRPM2 KO mice. Cells were perfused with either 300  $\mu$ M ADPR (green symbols,  $n = 7$ ), 300  $\mu$ M ADPR plus 1 mM AMP (blue symbols,  $n = 9$ ) or 3 mM ADPR (black symbols,  $n = 6$ ). TRPM2 KO cells were perfused with 1 mM ADPR (red symbols,  $n = 6$ ). Current amplitudes were assessed as described in Fig. 2A. Error bars indicate SEM. **(B)** Typical current-voltage ( $I$ - $V$ ) relationship of currents evoked by 1 mM ADPR (black trace), 300  $\mu$ M ADPR (green trace), or 300  $\mu$ M ADPR + 1 mM AMP (blue trace) taken from representative cells and recorded 100 s into the experiment. **(C)** Dose-response behavior of TRPM2 currents in mouse  $\beta$  cells as a function of internal ADPR concentration. Current amplitudes were measured at  $-80$  mV, averaged, normalized to cell size, and plotted versus the respective ADPR concentration ( $n = 5$  to 7). A dose-response fit to the data resulted in an  $EC_{50}$  value of 360  $\mu$ M

with a Hill coefficient of 1. **(D)** Average  $Ca^{2+}$  signals measured in intact fura-2-AM-loaded mouse  $\beta$  cells in response to increasing concentrations of extracellular ADPR and in the absence of extracellular  $Ca^{2+}$  [1  $\mu$ M (red trace,  $n = 4$ ), 10  $\mu$ M (blue trace,  $n = 5$ ), 30  $\mu$ M (green trace,  $n = 6$ ), 100  $\mu$ M (black trace,  $n = 6$ )]. Start of application indicated by black arrow. **(E)** Average  $Ca^{2+}$  signals measured in intact fura-2-AM-loaded mouse  $\beta$  cells in response to application of 200  $\mu$ M ADPR in the absence of extracellular  $Ca^{2+}$  and in the presence of either 100  $\mu$ M suramin (red trace,  $n = 6$ ) or 1  $\mu$ M CGS-15943 (black trace,  $n = 8$ ) in the external solution. **(F)** Balanced fura-2 experiments showing average  $Ca^{2+}$  signals in whole-cell patch-clamped C57BL/6 mouse pancreatic  $\beta$  cells (black trace,  $n = 7$ ) or  $\beta$  cells isolated from TRPM2 KO mice (red trace,  $n = 10$ ) preloaded with fura-2-AM. Whole-cell break-in indicated by red arrow. Cells were kept in 0  $Ca^{2+}$  external solution and perfused with internal solution containing 300  $\mu$ M ADPR and 200  $\mu$ M fura-2. The gray traces show two representative responses measured in individual wild-type (WT) cells.

NAD<sup>+</sup> and cADPR triggered Ca<sup>2+</sup> release transients in INS-1 cells, although cADPR did so more potently and effectively than NAD<sup>+</sup> (Fig. 6B). The threshold concentration for cADPR was ~10 μM (fig. S2A) and for NAD<sup>+</sup> ~30 μM (fig. S2C), just 10 to 30 times higher than that of ADPR. Like ADPR, both cADPR and NAD<sup>+</sup> effects were mediated through P2Y and adenosine receptors, because the combined suppression of these receptors by suramin and CGS-15943 completely blocked the response (figs. S2B and S2C, respectively). cADPR showed a pharmacological profile similar to that of ADPR, because suramin was more effective than CGS-15943 at suppressing the response to cADPR (fig. S2B). However, cADPR, even at 100 μM, failed to release Ca<sup>2+</sup> in the presence of the ADPR antagonist 8-bromo-ADPR [8-Br-ADPR; 100 μM, fig. S3; (26)].

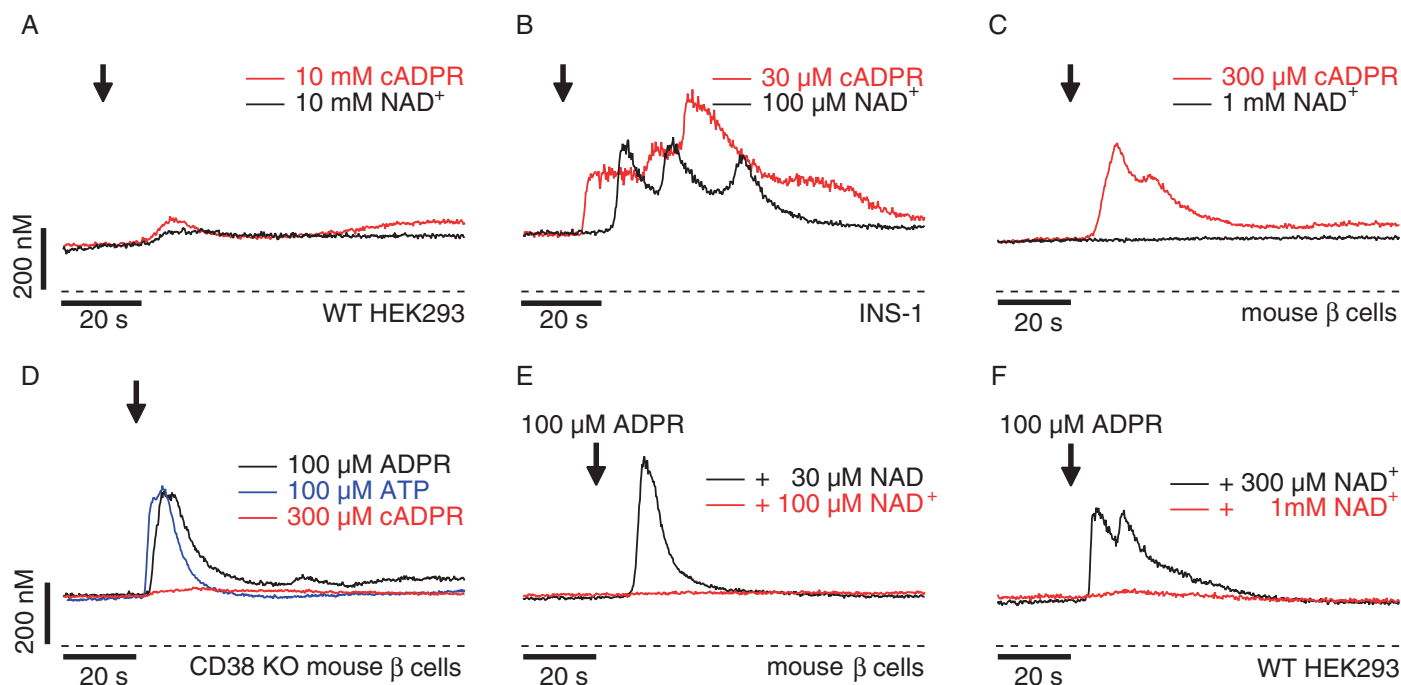
In primary β cells, extracellular NAD<sup>+</sup> did not elicit any response even at 1 mM (Fig. 6C). However, cADPR at 300 μM produced clear Ca<sup>2+</sup>-release transients when applied to intact cells (Fig. 6C). Because mouse β cells express CD38 (27) and cADPR acts as a substrate of the ectoenzyme CD38 to produce ADPR (10), we used a CD38 knockout mouse (28) to test whether the efficacy of cADPR relied on the presence of this enzyme. This was indeed the case; CD38-deficient β cells did not respond to cADPR, but retained responsiveness to ADPR (Fig. 6D). CD38-deficient β cells also retained functional P2Y receptors as evidenced by the Ca<sup>2+</sup>-release transient

induced by 100 μM ATP (Fig. 6D), suggesting that responsiveness to extracellular cADPR indeed requires CD38. Together, these data indicate that ADPR, and not cADPR, represents the primary P2Y receptor agonist.

We were puzzled by the lack of effect of NAD<sup>+</sup>, which is the major substrate for ADPR production by CD38 (10). This lack of effect could be explained if NAD<sup>+</sup> acts as a competitive inhibitor to ADPR-induced Ca<sup>2+</sup> release in mouse β cells. Indeed, addition of 100 μM NAD<sup>+</sup> in combination with 100 μM ADPR (1:1 ratio) to intact mouse β cells (Fig. 6E) completely suppressed the Ca<sup>2+</sup> release normally induced by 100 μM ADPR (Figs. 5, D and E, and 6D). However, a robust Ca<sup>2+</sup> signal was apparent with a NAD<sup>+</sup> concentration of 30 μM (Fig. 6E). Thus, although neither NAD<sup>+</sup> nor cADPR act as P2Y agonists, NAD<sup>+</sup> seems to act as a competitive inhibitor of ADPR at P2Y receptors in mouse β cells. NAD<sup>+</sup> also inhibited ADPR-induced Ca<sup>2+</sup> release in HEK293 cells, albeit with lower potency than in primary β cells (Fig. 6F). Although 100 μM ADPR combined with 300 μM NAD<sup>+</sup> still elicited Ca<sup>2+</sup> release, release was abolished by 1 mM NAD<sup>+</sup>, a 1:10 ratio of the two compounds.

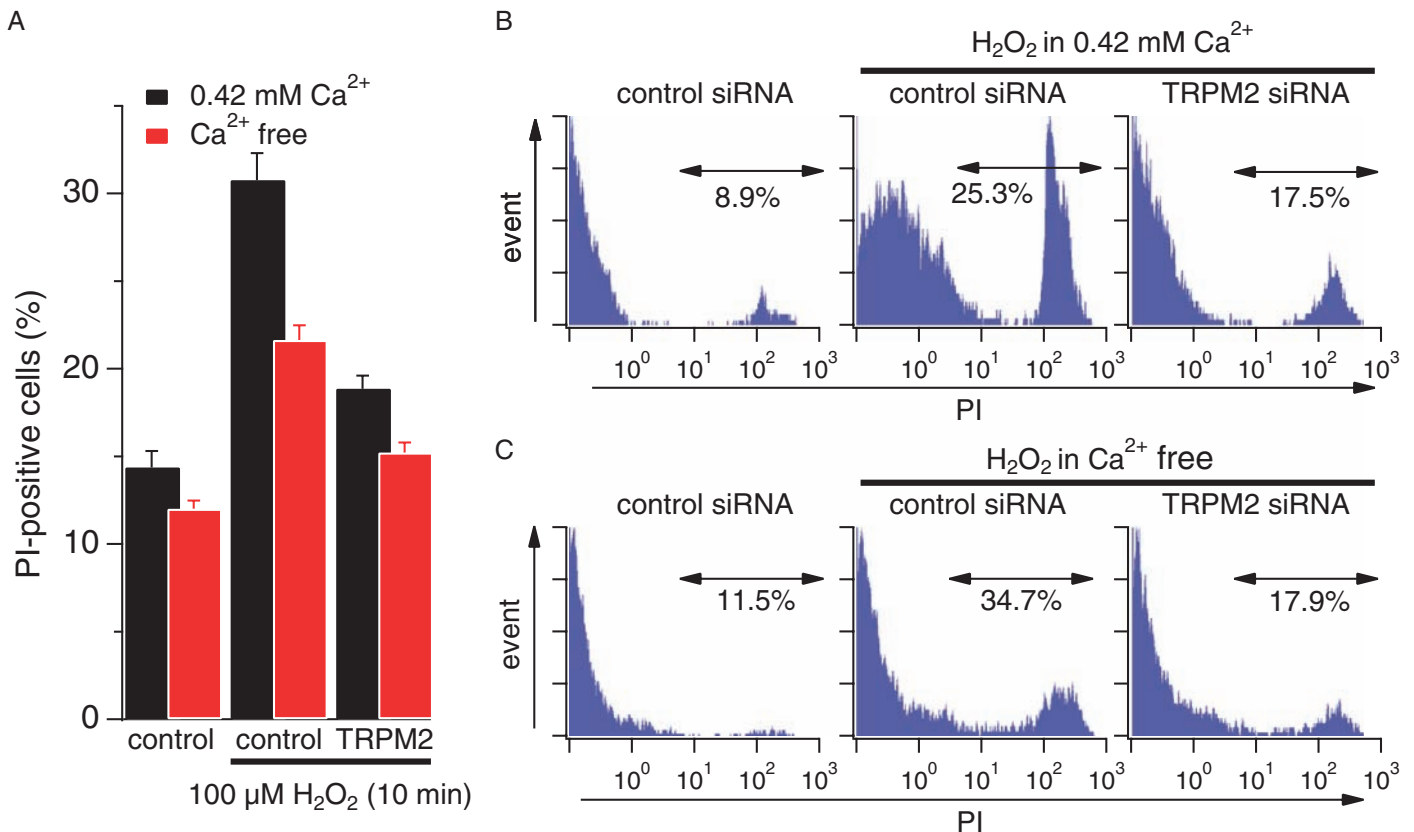
### Intracellular ADPR elicits Ca<sup>2+</sup> release in primary mouse β cells

Next, we investigated whether TRPM2 could mediate Ca<sup>2+</sup> release from intracellular stores of primary β cells. Figure 5F shows that intracellular



**Fig. 6.** The ectoenzyme CD38 is key in cADPR-induced Ca<sup>2+</sup> signals. (A) Average Ca<sup>2+</sup> signals in intact wild-type HEK293 cells in response to extracellular application of 10 mM cADPR (red trace,  $n = 8$ ) or 10 mM NAD<sup>+</sup> (black trace,  $n = 6$ ) in the absence of extracellular Ca<sup>2+</sup>. Application and fura-2-AM loading as described in Fig. 1A. (B) Average Ca<sup>2+</sup> signals in intact fura-2-AM-loaded INS-1 cells in response to 30 μM external cADPR (red trace,  $n = 5$ ) or 100 μM NAD<sup>+</sup> (black trace,  $n = 10$ ) in the absence of extracellular Ca<sup>2+</sup>. Application start is indicated by the arrow. (C) Average Ca<sup>2+</sup> signals measured in intact fura-2-AM-loaded primary mouse β cells in response to external application of either 300 μM cADPR (red trace,  $n = 6$ ) or 1 mM NAD<sup>+</sup> (black trace,  $n = 8$ ). (D) Average Ca<sup>2+</sup> signals measured in intact fura-2-AM-loaded pancreatic

β cells isolated from CD38 knockout mice (28) in response to external application of either 100 μM ADPR (black trace,  $n = 4$ ), 100 μM ATP (blue trace,  $n = 8$ ), or 300 μM cADPR (red trace,  $n = 20$ ). (E) Average Ca<sup>2+</sup> signals measured in intact fura-2-AM-loaded mouse pancreatic β cells. As indicated by the arrow, 100 μM ADPR was co-applied with either 30 μM NAD<sup>+</sup> (black trace,  $n = 6$ ) or 100 μM NAD<sup>+</sup> (red trace,  $n = 6$ ) in a 0 Ca<sup>2+</sup> solution. (F) Average Ca<sup>2+</sup> signals measured in wild-type HEK293 cells in response to application of extracellular ADPR (100 μM) in the presence of either 1 mM NAD<sup>+</sup> (red trace,  $n = 9$ ) or 300 μM NAD<sup>+</sup> (black trace,  $n = 7$ ) and in the absence of extracellular Ca<sup>2+</sup>. Application started as indicated by the arrow and was maintained throughout the experiment.



**Fig. 7.** TRPM2-mediated Ca<sup>2+</sup> release induces cell death under oxidative stress. **(A)** Average values for percent of PI-positive cells. INS-1 cells transfected with control (GAPDH) or TRPM2 siRNA were treated with 100  $\mu$ M H<sub>2</sub>O<sub>2</sub> for 10 min either in RPMI 1640 medium that contained 0.42 mM Ca<sup>2+</sup> (black bars,  $n = 5$ ) or was Ca<sup>2+</sup>-free (red bars,  $n = 11$ ). PI-positive cells were analyzed by flow cytometry. Data points are mean  $\pm$  SEM. Comparing H<sub>2</sub>O<sub>2</sub>-

treated and -untreated control (GAPDH) cells, H<sub>2</sub>O<sub>2</sub>-treated control (GAPDH) and TRPM2 siRNA cells, or untreated control (GAPDH) cells with H<sub>2</sub>O<sub>2</sub>-treated TRPM2 siRNA cells showed a statistical significance of  $P < 0.001$  in each case, both in the presence and absence of extracellular Ca<sup>2+</sup>. **(B)** Representative PI profile of cells tested in the presence of extracellular Ca<sup>2+</sup>. **(C)** Representative PI profile of cells tested in the absence of extracellular Ca<sup>2+</sup>.

perfusion of mouse  $\beta$  cells with 300  $\mu$ M ADPR evoked Ca<sup>2+</sup> release transients. Unlike the IP<sub>3</sub>-mediated Ca<sup>2+</sup> oscillations elicited by extracellular ADPR (Fig. 5D), internally applied ADPR typically gave rise to a single transient both in INS-1 cells and primary  $\beta$  cells (fig. S4D). However, increasing internal ADPR concentrations shortened the delay of this Ca<sup>2+</sup> transient and enhanced the overall change in peak amplitude of Ca<sup>2+</sup> release in INS-1 cells (fig. S4F). We further confirmed that the responses mediated by intracellular ADPR involved TRPM2 by investigating mouse primary  $\beta$  cells isolated from TRPM2 knockout mice (TRPM2 KO) (29). As illustrated in Fig. 5A, even 1 mM intracellular ADPR failed to evoke currents in these cells, confirming the absence of functional TRPM2 channels in the plasma membrane. Furthermore, Ca<sup>2+</sup>-release signals were completely absent in TRPM2 KO  $\beta$  cells perfused with 1 mM ADPR in the absence of extracellular Ca<sup>2+</sup> (Fig. 5F;  $P < 0.026$  as assessed by change in peak Ca<sup>2+</sup> release, table S1).

### Lysosomal Ca<sup>2+</sup> release through TRPM2 contributes to apoptosis in INS-1 $\beta$ cells

To examine the possible physiological function of TRPM2-mediated Ca<sup>2+</sup> release in pancreatic  $\beta$  cells, we tested INS-1 cells for H<sub>2</sub>O<sub>2</sub>-induced susceptibility to cell death assessed by analysis of propidium iodide (PI) stain-

ing in flow cytometry. Unfortunately, any molecular knockdown of TRPM2 would affect the expression of this channel in both plasma membrane and lysosomal compartments. We therefore compared the effect of TRPM2 on H<sub>2</sub>O<sub>2</sub>-induced cell death in the presence and absence of extracellular Ca<sup>2+</sup>. The latter abolishes Ca<sup>2+</sup> influx through TRPM2 but leaves internal Ca<sup>2+</sup> release through this channel intact, representing a functional knockout of plasma membrane TRPM2. These data were compared to results obtained from cells treated with TRPM2-specific siRNA, thereby isolating the severity of cell death linked to TRPM2-mediated Ca<sup>2+</sup> release. Exposure to 100  $\mu$ M H<sub>2</sub>O<sub>2</sub> significantly enhanced cell death in INS-1 cells treated with control siRNA [directed against glyceraldehyde-3-phosphate dehydrogenase (GAPDH)] in the presence of extracellular Ca<sup>2+</sup> ( $P < 0.001$ ). However, cells with suppressed TRPM2 expression were 72% less affected by H<sub>2</sub>O<sub>2</sub>-induced cell death (Fig. 7, A and B;  $P < 0.001$ ). As Fig. 7A further illustrates, H<sub>2</sub>O<sub>2</sub> was able to induce significant cell death in control siRNA (GAPDH)-treated INS-1 cells even in the absence of extracellular Ca<sup>2+</sup> ( $P < 0.001$ ), albeit with reduced severity. However, this was still linked to TRPM2 expression, because H<sub>2</sub>O<sub>2</sub>-induced cell death in the absence of extracellular Ca<sup>2+</sup> was 68% further reduced (Fig. 7, A and C;  $P < 0.001$ ) in cells transfected with TRPM2 siRNA. This indicates that not only Ca<sup>2+</sup> influx through plasma membrane TRPM2 but



also TRPM2-dependent lysosomal  $\text{Ca}^{2+}$  release plays a critical role in  $\text{H}_2\text{O}_2$ -mediated  $\beta$  cell death.

## DISCUSSION

We here establish that TRPM2, in addition to its known role as a plasma membrane-resident  $\text{Ca}^{2+}$ -influx channel, can also function as an intracellular  $\text{Ca}^{2+}$ -release channel in lysosomes of pancreatic  $\beta$  cells. Intracellular ADPR is capable of activating these TRPM2 functions, and both pathways contribute to  $\text{H}_2\text{O}_2$ -induced  $\beta$  cell death. Independent of its intracellular agonistic action on TRPM2, extracellular ADPR acts as a primary P2Y and adenosine receptor agonist, resulting in  $\text{IP}_3$  formation and  $\text{Ca}^{2+}$  release from the ER.  $\text{NAD}^+$  and cADPR are ineffective as direct agonists of either P2Y or adenosine receptors, but can indirectly cause  $\text{IP}_3$ -dependent  $\text{Ca}^{2+}$  release through metabolic conversion to ADPR by the ectoenzyme CD38. Although  $\text{NAD}^+$  does not act as a P2Y receptor agonist in the cells investigated here, it functions as a competitive antagonist of ADPR at the P2Y subtypes expressed in HEK293 and primary mouse  $\beta$  cells.

TRPM2 has been extensively characterized as a plasma membrane ion channel that is specifically activated by ADPR (1–3). TRPM2 sensitivity to ADPR is regulated by facilitatory cofactors such as cytosolic  $\text{Ca}^{2+}$  or production of cADPR and NAADP (5, 7). These molecules act synergistically and sensitize TRPM2 to ADPR so that, for example, the apparent  $\text{EC}_{50}$  of ADPR of  $\sim 100 \mu\text{M}$  in HEK293 cells is reduced to  $\text{EC}_{50}$  concentrations as low as 90 nM in the presence of cADPR (5). In addition, different cellular systems exhibit different ADPR sensitivity of TRPM2 for unknown reasons (1, 5, 7). Although high micromolar concentrations of agonist may occur during pathophysiological events, it is possible or even likely that under physiological conditions, synergistic events may facilitate TRPM2 activation.

The present study now adds previously unrecognized facets to the function of TRPM2 and its natural ligands by establishing TRPM2 as an intracellular  $\text{Ca}^{2+}$ -release channel and showing that ADPR and its precursors  $\text{NAD}^+$  and cADPR also exhibit extracellular activity as receptor agonists and antagonists. Although ADPR has been shown to activate  $\text{Ca}^{2+}$  signaling through  $\text{IP}_3$ -producing receptors, the specific receptor species involved has not yet been identified (13). Our results show that ADPR can activate P2Y receptors in the three cell types investigated in this study (HEK293, INS-1, and primary mouse  $\beta$  cells), as well as adenosine receptors in INS-1 cells. However, ADPR was about two orders of magnitude more potent in  $\beta$  cells than in HEK293 cells. Possible reasons for this greater sensitivity include species differences in P2Y sensitivity, different complements of P2Y receptor subtypes, or both. HEK293 cells mainly express P2Y subtypes 1, 2, and 4 (16), although a slightly differing P2Y receptor complement has also been reported for these cells (30). INS-1 cells express subtypes 1, 2, 4, 6, and 12 in similar amounts (16, 31). Thus, a specific P2Y receptor subtype complement, possibly involving subtypes 6 or 12 or both, might be responsible for the high-affinity response to ADPR in INS-1 cells.

ADPR can be produced extracellularly from its precursors  $\text{NAD}^+$  or cADPR through the action of the ectoenzyme CD38 (10). Two recent studies report that  $\text{NAD}^+$  itself may be an agonist for P2Y11 receptors in granulocytes (32) and P2Y1 receptors in visceral smooth muscle (33). However, even at high millimolar concentrations, neither  $\text{NAD}^+$  nor cADPR produced  $\text{Ca}^{2+}$  signals in HEK293 (Fig. 6A). This suggests that these molecules are not effective agonists for the P2Y receptor subtypes endogenously expressed in HEK293 cells or that these cells may not express enough CD38 to produce substantial amounts of ADPR from these precursors.

In INS-1 cells, however,  $\text{NAD}^+$  and cADPR caused  $\text{Ca}^{2+}$  release with an  $\sim 10$  times increased efficiency compared to that of HEK293 cells. This is either due to a genuine agonistic action of these compounds on cell sur-

face receptors or caused by exogenous metabolic conversion to ADPR through CD38, which is abundant in  $\beta$  cells (34). Although CD38 accepts both  $\text{NAD}^+$  and cADPR as substrates and converts them to the common product ADPR, it appears to process  $\text{NAD}^+$  more efficiently than cADPR (35–37). cADPR induced  $\text{Ca}^{2+}$  release in both INS-1 and primary  $\beta$  cells, albeit 10 times less potently than ADPR. Its ability to do so can be linked to metabolic conversion of cADPR to ADPR through CD38; cADPR was ineffective in eliciting  $\text{Ca}^{2+}$  release in primary  $\beta$  cells of transgenic mice deficient in CD38, despite an intact P2Y pathway (Fig. 6D).  $\text{NAD}^+$  was ineffective in triggering  $\text{Ca}^{2+}$  release in primary  $\beta$  cells, likely because it acted as a competitive antagonist to ADPR, suppressing specific P2Y receptor-mediated  $\text{Ca}^{2+}$  signals in primary  $\beta$  cells (Fig. 6E) and HEK293 cells (Fig. 6F). Although  $\text{NAD}^+$  may well be converted to ADPR by CD38, the action of ADPR is prevented by the concomitant inhibition of P2Y receptors by  $\text{NAD}^+$  itself. Nevertheless, extracellular  $\text{NAD}^+$  does cause  $\text{Ca}^{2+}$  release in the INS-1 cell line. Although this effect could be due to the specific P2Y subtype expression pattern in these cells or, alternatively, caused by direct adenosine receptor activation, our data are most consistent with a metabolic conversion of  $\text{NAD}^+$  to ADPR, which activates adenosine receptors present in INS-1 cells; not only is there a substantial delay in the  $\text{Ca}^{2+}$  release response to  $\text{NAD}^+$  application (Fig. 6B), the signal itself is smaller than that elicited by ADPR (Fig. 2D). Thus, the precise P2Y receptor subtype composition of a cell, as well as the expression of adenosine receptors and CD38, would determine the strength of the resulting  $\text{Ca}^{2+}$  signal elicited in the presence of ADPR and  $\text{NAD}^+$ . Given that the extracellular effects of ADPR are entirely mediated through membrane receptors, it appears that ADPR is not transported across the plasma membrane to an extent necessary to activate TRPM2.

The present study identifies TRPM2 as an intracellular  $\text{Ca}^{2+}$ -release channel localized in lysosomal compartments. Our data reveal that intracellular ADPR causes  $\text{Ca}^{2+}$  release only in TRPM2-expressing cells, but not in wild-type HEK293 cells, and that this  $\text{Ca}^{2+}$  release is sensitive to the TRPM2-specific antagonist AMP. Furthermore, endogenously expressed TRPM2 also functions as a  $\text{Ca}^{2+}$ -release channel in rat INS-1 and primary mouse  $\beta$  cells, because intracellular ADPR-induced  $\text{Ca}^{2+}$  release is reduced in siRNA experiments suppressing TRPM2 in INS-1 cells and absent in  $\beta$  cells isolated from TRPM2 knockout mice. Both immunofluorescence and functional data confirm the presence of TRPM2 predominantly in a lysosomal compartment rather than the ER. Accordingly, neither heparin nor ryanodine interfered with the  $\text{Ca}^{2+}$  release activity induced by intracellular ADPR, although the response to ADPR was somewhat blunted by ryanodine (Fig. 3B). This could mean that ryanodine at the relatively high concentration used here has nonspecific effects on TRPM2 channels or that ryanodine receptors may partially colocalize with TRPM2 in a store subcompartment, possibly indicating some cross talk between lysosomal and ER  $\text{Ca}^{2+}$  stores due to a small component of  $\text{Ca}^{2+}$ -induced  $\text{Ca}^{2+}$  release.

Oxidative stress due to production of ROS is thought to play a central role in  $\beta$  cell death and development of diabetes types 1 and type 2 (38, 39).  $\beta$  cells have only modest capacity for self-protection against ROS, including  $\text{H}_2\text{O}_2$ , because of their low expression of antioxidant enzymes, in particular glutathione peroxidase and catalase, which decompose  $\text{H}_2\text{O}_2$  (40). Exposure of MIN6  $\beta$  cells to  $\text{H}_2\text{O}_2$  has been reported to induce  $\text{Ca}^{2+}$ -dependent cell death involving both extracellular  $\text{Ca}^{2+}$  and release from  $\text{Ca}^{2+}$  stores (41). Although some reports have linked TRPM2 to  $\text{H}_2\text{O}_2$ -induced  $\text{Ca}^{2+}$  influx and cell death (42, 43), we here identify this channel as a novel mechanism in this process through its function as a lysosomal  $\text{Ca}^{2+}$ -release channel. Because release of  $\text{Ca}^{2+}$  from lysosomes is critical for the redistribution of phosphatidylserine (PS) from the inner plasma membrane leaflet to the cell surface (44), TRPM2-mediated  $\text{Ca}^{2+}$  release

may not only contribute to apoptosis itself, but additionally represent a crucial element for the externalization of PS, a key recognition ligand for the ultimate elimination of apoptotic cells.

In summary, TRPM2 and its agonist ADPR are multifunctional elements of the  $\beta$  cell  $\text{Ca}^{2+}$  signaling machinery. External ADPR functions as a first messenger for receptor-mediated  $\text{IP}_3$  signaling through P2Y and adenosine receptors and activation of TRPM2 by internal ADPR contributes both to  $\text{Ca}^{2+}$  entry across the plasma membrane and  $\text{Ca}^{2+}$  release from lysosomes, affecting  $\text{Ca}^{2+}$ -dependent apoptosis.

## MATERIALS AND METHODS

### Cells

Full-length TRPM2 complementary DNA was cloned into a modified version of the pCDNA4/TO vector (Invitrogen) with an N-terminal Flag epitope tag and electroporated into HEK293 cells previously transfected with the pCDNA6/TR construct for Tet repressor expression as described (1). Pancreatic  $\beta$  cells were isolated from C57BL/6 wild-type or CD38 knockout mice as described (45) and as approved by the Institutional Animal Care and Use Committee, University of Hawaii, and the Animal Care Committee, The Queen's Medical Center.

### Electrophysiology

External solution contained (in mM) 140 NaCl, 2.8 KCl, 1  $\text{CaCl}_2$ , 2  $\text{MgCl}_2$ , 10 glucose, 10 HEPES-NaOH (pH 7.2 adjusted with NaOH). Internal solution contained (in mM) 140 cesium glutamate for INS-1 and primary  $\beta$  cells, 140 potassium glutamate for HEK293, 8 NaCl, 1  $\text{MgCl}_2$ , 10 Hepes-Cs/KOH. ADPR was added as appropriate. Reagents were from Sigma-Aldrich except Suramin (Fluka). 8-Br-ADPR was synthesized as described (26). Relative purity of 8-Br-ADPR was assessed by HPLC analyses and found greater than 95%. No traceable 8-Br-NAD<sup>+</sup> or 8-Br-cADPR contaminants were detected (46). Patch-clamp experiments were performed as described (1). Error bars indicate SEM with *n* determinations.

### Fluorescence measurements

Fluorescence signals were sampled at a rate of 5 Hz with a photomultiplier-based system using a monochromatic light source (TILL Photonics, Gräfelfing, Germany). Emission was detected with a photomultiplier whose analog signals were sampled by a digital-analog interface (ITC-16, Instrutech, New York) and processed by X-Chart software (HEKA, Lambrecht, Germany). Fluorescence ratios were converted into free intracellular  $\text{Ca}^{2+}$  concentration based on calibration parameters derived from patch-clamp experiments with calibrated  $\text{Ca}^{2+}$  concentrations. Three different kinds of fluorescence experiments were performed. In experiments combining patch-clamp and fluorescence experiments, cells were perfused with standard intracellular pipette solution containing 200  $\mu\text{M}$  fura-2. Balanced fura-2 experiments were performed by preloading cells with fura-2-AM at 5  $\mu\text{M}$  for 30 minutes. In the subsequent whole-cell patch clamp experiments 200  $\mu\text{M}$  fura-2 had been added to the standard internal solution to assure continuous fura-2 signals. For intact-cell  $\text{Ca}^{2+}$  measurements, cells were loaded with 5  $\mu\text{M}$  fura-2-AM for 30 minutes.

For detailed descriptions, see Supplementary Materials and Methods.

## SUPPLEMENTARY MATERIALS

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Materials and Methods

Fig. S1. Thapsigargin prevents  $\text{Ca}^{2+}$  release induced by extracellular ADPR.

Fig. S2. Extracellular cADPR and NAD<sup>+</sup> evoke  $\text{Ca}^{2+}$  signals in INS-1  $\beta$  cells.

Fig. S3. 8-Br-ADPR application inhibits  $\text{Ca}^{2+}$  release by cADPR.

Fig. S4. Statistical analysis of  $\text{Ca}^{2+}$  signals induced by external or internal ADPR in  $\beta$  cells.

Table S1. Statistical analysis of agonist-induced  $\text{Ca}^{2+}$  signals in TRPM2 HEK and  $\beta$  cells.

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